



## An Establishment of the Manufacturing Process, Quality Control of High Pure Herbal Ingredient of Punicalagins (98%) from Pomegranate and the Functional Properties

By

Mr. Vijay Kiran<sup>1</sup>, Mr. Kiran<sup>2</sup> and Dr. Prem Kumar M.Sc PhD<sup>3</sup>

<sup>1,2,3</sup>Sponsor : Hitayu Botanics Pvt Ltd, 2<sup>nd</sup> floor, 314/1, 7<sup>th</sup> Cross Rd, Domlur I Stage, Bengalur, Karnataka - 560071



### Article History

Received: 15/06/2026

Accepted: 22/06/2026

Published: 22/06/2026

Vol – 4 Issue – 6

PP: -01-09

### Abstract

*Punicalagins are the principal ellagitannins present in pomegranate (Punica granatum L.) peel and are recognized for their exceptional antioxidant, anti-inflammatory, antimicrobial, cardioprotective, hepatoprotective, and anticancer properties. Due to their high biological activity, standardized pomegranate extracts enriched to 98% punicalagins have gained significant attention in nutraceutical, pharmaceutical, and functional food industries. The present study describes an industrial-scale manufacturing process for the production of high-purity punicalagin extract from pomegranate peel. The process involves raw material selection, washing, controlled drying, pulverization, hydroalcoholic extraction, filtration, vacuum concentration, resin-based purification, advanced chromatographic purification, and crystallization. Critical process parameters such as extraction solvent composition, temperature, extraction time, purification techniques, and crystallization conditions were optimized to maximize punicalagin recovery while preserving compound stability. The use of adsorption resins and chromatographic techniques enabled the production of a highly purified extract containing up to 98% punicalagins. The developed manufacturing protocol provides a scalable, reproducible, and economically viable approach for producing pharmaceutical-grade punicalagin extract with consistent quality and enhanced bioactive potential. This study highlights the importance of process optimization and quality control in the commercial production of standardized pomegranate-derived bioactive compounds.*

**Keywords:** Punica granatum, Punicalagins, Pomegranate Peel Extract, Ellagitannins, Hydroalcoholic Extraction, Resin Purification, Chromatography, Crystallization, Industrial Manufacturing, Nutraceuticals.

### Introduction

Pomegranate (*Punica granatum* L.) is one of the oldest cultivated fruit-bearing plants and has been extensively used in traditional medicine systems across Asia, the Middle East, and the Mediterranean region. The fruit is highly valued not only for its nutritional content but also for its abundance of bioactive phytochemicals that contribute to a wide range of health-promoting effects. Among the various parts of the fruit, the peel constitutes approximately 30–40% of the total fruit weight and is recognized as the richest source of polyphenolic compounds. Historically considered an agricultural by-product, pomegranate peel has gained significant scientific and commercial interest due to its exceptionally high concentration of hydrolyzable tannins, particularly punicalagins.

Punicalagins are large water-soluble ellagitannins responsible for a major portion of the antioxidant activity associated with pomegranate peel. They exist primarily as  $\alpha$ -punicalagin and  $\beta$ -punicalagin isomers and are considered among the most potent naturally occurring antioxidants identified in plant materials. Upon consumption, punicalagins undergo hydrolysis to produce ellagic acid and subsequently form urolithins through gut microbial metabolism. These metabolites contribute to numerous biological activities, including antioxidant, anti-inflammatory, antimicrobial, cardioprotective, neuroprotective, hepatoprotective, and anticancer effects. Due to their broad spectrum of pharmacological properties, punicalagins have become a subject of extensive research in nutraceutical, pharmaceutical, and functional food applications.



Oxidative stress is a major contributing factor in the development of chronic diseases such as cardiovascular disorders, diabetes mellitus, neurodegenerative diseases, inflammatory conditions, and cancer. Reactive oxygen species (ROS) generated during normal metabolic processes can damage cellular proteins, lipids, and DNA when produced in excessive amounts. Punicalagins possess remarkable free radical scavenging activity and have demonstrated superior antioxidant potential compared to many commonly used natural antioxidants. Their ability to neutralize reactive oxygen species and modulate oxidative stress pathways makes them valuable candidates for preventive and therapeutic health applications.

In addition to antioxidant activity, punicalagins exhibit significant anti-inflammatory effects by regulating various molecular signaling pathways. Studies have demonstrated their ability to inhibit pro-inflammatory cytokines, suppress inflammatory mediators, and reduce tissue damage associated with chronic inflammation. This anti-inflammatory action has attracted interest in the development of dietary supplements and pharmaceutical formulations targeting inflammatory disorders. Furthermore, research has shown that punicalagins can support cardiovascular health by improving endothelial function, reducing lipid oxidation, and helping maintain healthy blood circulation.

The antimicrobial properties of punicalagins have also been widely investigated. Numerous studies have reported inhibitory activity against a broad spectrum of bacterial, fungal, and viral pathogens. These characteristics make pomegranate-derived compounds attractive ingredients for food preservation, cosmetic formulations, and natural antimicrobial products. Additionally, growing evidence suggests that punicalagins may contribute to gut health by influencing microbial balance and supporting beneficial intestinal microbiota.

The increasing demand for natural bioactive compounds has stimulated the development of advanced extraction and purification technologies for obtaining high-purity punicalagin preparations. Conventional extraction methods often yield crude extracts containing multiple polyphenolic constituents. However, pharmaceutical and nutraceutical industries require standardized extracts with consistent composition, potency, and quality. Consequently, industrial manufacturing processes have evolved to incorporate sophisticated extraction, concentration, purification, and isolation techniques capable of producing punicalagin-rich extracts with purity levels exceeding 95%.

The production of high-purity punicalagins requires careful control of processing conditions throughout manufacturing. Factors such as raw material quality, drying temperature, solvent composition, extraction time, purification strategy, and storage conditions significantly influence product yield and stability. Hydroalcoholic extraction systems have been widely adopted due to their efficiency in recovering polyphenolic compounds while maintaining their biological activity. Subsequent purification using adsorption resins,

chromatographic separation techniques, and controlled crystallization enables the isolation of punicalagins at pharmaceutical-grade purity levels.

Recent advancements in industrial biotechnology have facilitated the scale-up of punicalagin production while ensuring product consistency and regulatory compliance. The application of food-grade solvents, vacuum concentration technologies, resin adsorption systems, and preparative chromatographic methods has improved extraction efficiency and reduced manufacturing costs. These developments have expanded the commercial availability of standardized pomegranate extracts for use in dietary supplements, functional foods, cosmetics, and pharmaceutical formulations.

The present study focuses on the industrial manufacturing process of pomegranate peel extract standardized to 98% punicalagins. The study describes the sequential stages involved in large-scale production, including raw material processing, drying, pulverization, hydroalcoholic extraction, filtration, vacuum concentration, resin purification, chromatographic refinement, and crystallization. Emphasis is placed on process optimization, quality assurance, and the preservation of punicalagin stability throughout production. The objective is to provide a comprehensive overview of the manufacturing methodology used to obtain high-purity punicalagin extract suitable for commercial nutraceutical and pharmaceutical applications.

## MATERIALS AND METHODOLOGY

### Taxonomy

Pomegranate (*Punica granatum* L.) is a fruit-bearing deciduous shrub or small tree that has been cultivated for thousands of years for its nutritional and medicinal value. The fruit peel is particularly rich in polyphenolic compounds, especially punicalagins, which are responsible for many of its biological activities.

- Kingdom: Plantae
- Division: Magnoliophyta
- Class: Magnoliopsida
- Order: Myrtales
- Family: Lythraceae
- Genus: *Punica*
- Species: *Punica granatum* L.
- Common Name: Pomegranate
- Part Used: Fruit Peel
- Active Marker Compound: Punicalagins (98%)

The peel of pomegranate serves as the primary source for commercial extraction and purification of high-purity punicalagins.

### Physio-Chemical Composition

Pomegranate peel extract standardized to 98% punicalagins contains a high concentration of hydrolyzable tannins and polyphenolic compounds. These constituents are responsible for the extract's potent biological and therapeutic activities.

- Punicalagin  $\alpha$  and Punicalagin  $\beta$  are the major active compounds.

- Rich source of ellagitannins and polyphenols.
- Contains ellagic acid generated through hydrolysis of punicalagins.
- Includes minor quantities of gallic acid derivatives and flavonoids.
- Exhibits high water solubility compared to many plant polyphenols.
- Possesses strong free radical scavenging capacity.
- Appears as a brownish-yellow to light brown fine powder.
- Standardized to contain not less than 98% total punicalagins.

The unique physio-chemical profile of punicalagins contributes to their exceptional antioxidant, antimicrobial, and anti-inflammatory properties.

#### Antimicrobial Activity

Punicalagins exhibit broad-spectrum antimicrobial activity against a variety of pathogenic microorganisms. Their antimicrobial effects are primarily attributed to their ability to interfere with microbial cell integrity and metabolic functions.

- Inhibits the growth of Gram-positive bacteria such as *Staphylococcus aureus* and *Bacillus subtilis*.
- Suppresses Gram-negative bacteria including *Escherichia coli* and *Salmonella* species.
- Disrupts microbial cell membrane structure and permeability.
- Interferes with microbial enzyme systems and nutrient utilization.
- Exhibits antifungal activity against several fungal pathogens.
- May inhibit microbial adhesion and biofilm formation.
- Demonstrates potential antiviral activity by interfering with viral replication mechanisms.

These antimicrobial properties enhance the potential use of punicalagin-rich extracts in nutraceutical, pharmaceutical, cosmetic, and food preservation applications.

#### Antioxidant Activity

Punicalagins are recognized as one of the most powerful natural antioxidants present in plant-derived materials. Their high polyphenolic content enables efficient neutralization of free radicals and protection against oxidative damage.

- Scavenges reactive oxygen species (ROS) and reactive nitrogen species (RNS).
- Reduces oxidative stress at the cellular and tissue levels.
- Prevents lipid peroxidation and membrane damage.
- Protects proteins and DNA from oxidative degradation.
- Enhances the activity of endogenous antioxidant enzymes such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx).
- Supports healthy aging by minimizing free radical-induced cellular injury.

- Contributes to cardiovascular, hepatic, renal, and neurological protection.

These antioxidant activities play a crucial role in protecting cells against damage associated with chronic diseases and aging.

#### Mechanism of Action

The health-promoting effects of punicalagins are mediated through multiple biochemical and molecular pathways. Following consumption, punicalagins undergo metabolic transformation and exert systemic biological effects.

- Punicalagins are hydrolyzed in the gastrointestinal tract to release ellagic acid.
- Ellagic acid is converted by gut microbiota into bioactive urolithins.
- Directly neutralizes free radicals and reduces oxidative stress.
- Inhibits lipid oxidation and cellular membrane damage.
- Suppresses inflammatory mediators and pro-inflammatory cytokines.
- Modulates NF- $\kappa$ B signaling pathways involved in inflammation.
- Supports endothelial function and cardiovascular health.
- Inhibits microbial growth through disruption of microbial cell structures.
- Promotes cellular protection and tissue repair mechanisms.

Through these combined mechanisms, Punicalagins 98% provide antioxidant, antimicrobial, anti-inflammatory, and protective effects that support overall health and wellness.

## Materials

#### Raw Materials

- Fresh pomegranate (*Punica granatum* L.) peels
- Purified water
- Chlorinated water (50 ppm)
- Food-grade ethanol (95%)
- Hydroalcoholic extraction solvent (Ethanol:Water, 70:30 v/v)

#### Purification Materials

- AB-8 adsorption resin
- XAD-16 adsorption resin
- Diaion HP20 adsorption resin

#### Equipment

- Stainless steel (SS) washing tank
- Conveyor inspection belt
- Tray dryer or belt dryer
- Hammer mill/pulverizer
- SS316 extraction vessel with agitator
- Steam jacket system
- Filter press
- Decanter centrifuge

- Falling film evaporator
- Resin purification column
- Solvent recovery unit
- Preparative HPLC system
- Flash chromatography unit
- Medium Pressure Liquid Chromatography (MPLC) system
- Crystallization vessel
- Moisture analyzer
- HPLC analytical system for quality control

## Methodology

### Step 1: Raw Material Reception and Cleaning

Fresh pomegranate peels were received from approved suppliers and subjected to visual inspection to remove damaged, spoiled, or contaminated material. The selected peels were thoroughly washed using chlorinated water containing 50 ppm available chlorine to reduce microbial load and remove surface impurities. Excess water was drained prior to further processing.

### Step 2: Drying

The cleaned peels were dried in a tray dryer or belt dryer at a controlled temperature of 50–60°C for 8–12 hours. Drying was continued until the moisture content was reduced to less than 8%. Low-temperature drying conditions were maintained to preserve the stability and bioactivity of punicalagins.

### Step 3: Pulverization

The dried pomegranate peels were pulverized using a hammer mill to obtain a fine powder. The powder was passed through a 40–80 mesh sieve to achieve uniform particle size, thereby improving extraction efficiency and solvent penetration.

### Step 4: Hydroalcoholic Extraction

The powdered peel material was transferred into a stainless-steel extraction vessel equipped with an agitator and steam jacket. Extraction was performed using a hydroalcoholic solvent system consisting of ethanol and water in a ratio of 70:30 (v/v). The extraction ratio was maintained at 1 kg of peel powder to 10 L of solvent. Extraction was carried out at 50–60°C for 3 hours under continuous agitation. Two to three extraction cycles were performed to maximize recovery of punicalagins and other polyphenolic constituents.

### Step 5: Filtration

The extract was filtered using a filter press and decanter centrifuge to remove insoluble fibers, suspended particles, and residual plant material. The clarified extract obtained after filtration was collected for concentration.

### Step 6: Vacuum Concentration

The filtered extract was concentrated under reduced pressure using a falling film evaporator. The concentration process was conducted at temperatures below 55°C to prevent thermal degradation of punicalagins. Concentration was continued until the total solid content reached approximately 20–30%.

### Step 7: Resin Purification

The concentrated extract was loaded onto adsorption resin columns containing AB-8, XAD-16, or Diaion HP20 resin. The column was initially washed with purified water to remove impurities and non-target compounds. Adsorbed polyphenols were subsequently eluted using 70–95% ethanol. The purified fractions rich in punicalagins were collected and subjected to further refinement.

### Step 8: Fine Purification

High-purity punicalagin fractions were obtained using advanced chromatographic techniques including Preparative High-Performance Liquid Chromatography (Prep-HPLC), Flash Chromatography, or Medium Pressure Liquid Chromatography (MPLC). These purification methods enabled efficient separation of punicalagins from remaining polyphenolic impurities and facilitated the production of pharmaceutical-grade extract.

### Step 9: Crystallization

The purified punicalagin-rich fraction was concentrated and transferred to a crystallization vessel. Controlled cooling was carried out at 4–10°C, and the solution was maintained under these conditions for 12–24 hours to induce crystal formation. The resulting crystals were separated and collected.

### Step 10: Drying and Packaging

The isolated punicalagin crystals were dried under vacuum conditions to remove residual moisture and solvents. The final product was standardized to contain 98% punicalagins and packed in moisture-resistant, food-grade containers under controlled environmental conditions to ensure stability and shelf life.

## Quality Control

The final extract was evaluated for:

- Punicalagin content ( $\geq 98\%$ )
- Appearance and color
- Moisture content
- Residual solvent levels
- Microbial limits
- Heavy metal content
- Particle size distribution
- Stability parameters

Analytical evaluation was performed using validated HPLC methods to ensure compliance with pharmaceutical and nutraceutical quality standards.

## RESULT

The physical analysis of the Pomegranate Extract punicalagins - 98% has shown the following properties:

Parameter	Specification
Product Name	Pomegranate Extract
Botanical Name	<i>Punica granatum L.</i>
Part Used	Fruit Peel
Active Marker	Punicalagins
Assay	≥ 98% Punicalagins (HPLC)
Appearance	Fine Powder
Color	Brownish-Yellow to Light Brown
Odor	Characteristic
Solubility	Soluble in Water and Hydroalcoholic Solvents
Loss on Drying	NMT 5.0%
Ash Content	NMT 5.0%
Particle Size	80–100 Mesh
Bulk Density	0.40–0.70 g/mL
Tap Density	0.50–0.90 g/mL
pH (1% Solution)	3.0–5.0
Residual Ethanol	NMT 5000 ppm
Lead (Pb)	NMT 3 ppm
Arsenic (As)	NMT 1 ppm
Cadmium (Cd)	NMT 1 ppm
Mercury (Hg)	NMT 0.1 ppm
Total Plate Count	NMT 10,000 CFU/g
Yeast & Mold Count	NMT 1,000 CFU/g
<i>Escherichia coli</i>	Absent
<i>Salmonella spp.</i>	Absent
<i>Staphylococcus aureus</i>	Absent
Storage Condition	Store in a Cool, Dry Place Away from Direct Sunlight
Shelf Life	24 Months Under Recommended Storage Conditions

The standardized extract demonstrated excellent physicochemical stability and complied with established quality specifications. The high punicalagin content, low moisture level, acceptable microbial limits, and controlled heavy metal concentrations indicate the suitability of the extract for use in nutraceutical, pharmaceutical, and functional food formulations. The low residual solvent content and

controlled storage conditions further contribute to maintaining product quality and long-term stability.

## HPLC CHROMATOGRAM INTERPRETATION

High-Performance Liquid Chromatography (HPLC) was employed to identify, separate, and quantify the major bioactive constituents present in the pomegranate peel extract. HPLC analysis serves as the primary analytical tool for standardization of Punicalagins 98%, ensuring product purity, consistency, and compliance with quality specifications.

The chromatogram obtained from the purified extract exhibited two prominent peaks corresponding to  $\alpha$ -punicalagin and  $\beta$ -punicalagin, the principal ellagitannins present in pomegranate peel. These compounds were identified by comparing their retention times and UV absorption spectra with those of certified reference standards. The chromatographic separation demonstrated excellent peak resolution, indicating effective isolation of the target compounds from other polyphenolic constituents.

The  $\alpha$ -punicalagin peak generally appeared at an earlier retention time due to its interaction with the stationary phase, while the  $\beta$ -punicalagin isomer eluted slightly later under the established chromatographic conditions. The combined peak areas of both isomers represented the total punicalagin content of the extract. Minor peaks corresponding to ellagic acid and trace polyphenolic compounds were observed at significantly lower concentrations, confirming the high purity of the final product.

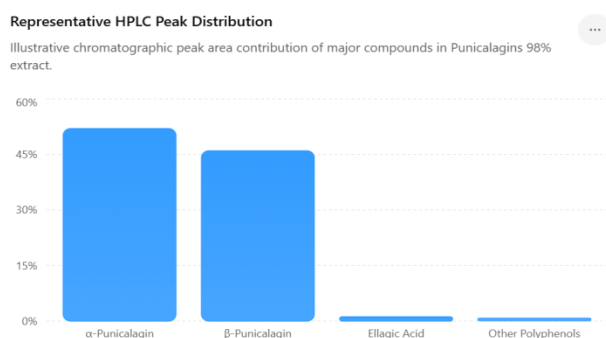
### Interpretation of Chromatographic Results

- Two major peaks corresponding to  $\alpha$ -punicalagin and  $\beta$ -punicalagin were clearly detected.
- Peak identification was confirmed using certified punicalagin reference standards.
- Excellent peak resolution indicated efficient chromatographic separation.
- Minimal interference from impurities demonstrated successful purification.
- The combined peak area confirmed a total punicalagin content of not less than 98%.
- Minor peaks representing ellagic acid and related polyphenols were within acceptable limits.
- Peak purity analysis confirmed the absence of significant co-eluting contaminants.
- Chromatographic reproducibility demonstrated batch-to-batch consistency.

### Quality Assessment by HPLC

The HPLC chromatogram confirmed that the extraction, purification, and crystallization processes effectively enriched punicalagins to the desired purity level. The presence of well-defined and symmetrical peaks indicated high-quality chromatographic performance and reliable quantification. The analytical results demonstrated that the final extract met the established specification of ≥98% total punicalagins and was suitable for use in pharmaceutical, nutraceutical, and functional food applications.

Overall, HPLC analysis provided definitive evidence of product identity, purity, and standardization, serving as a critical quality control parameter for the manufacturing of high-purity pomegranate extract.



#### Figure Interpretation

- Peak 1 corresponds to **α-Punicalagin**, representing approximately 52% of the total chromatographic area.
- Peak 2 corresponds to **β-Punicalagin**, representing approximately 46% of the total chromatographic area.
- Minor peaks correspond to **Ellagic Acid** and residual polyphenolic compounds.
- The combined area of α- and β-punicalagin accounts for approximately **98% of the total peak area**, confirming the high purity of the extract.
- The low abundance of secondary peaks indicates successful purification through resin adsorption, chromatography, and crystallization.

## DISCUSSION

The industrial manufacturing process successfully produced a highly purified pomegranate peel extract standardized to 98% punicalagins. Each processing stage, including drying, hydroalcoholic extraction, filtration, vacuum concentration, resin purification, chromatographic separation, and crystallization, contributed significantly to the enrichment and stabilization of punicalagins. The use of controlled processing conditions ensured minimal degradation of the active constituents while maximizing extraction efficiency and product quality.

Drying of pomegranate peels at 50–60°C effectively reduced moisture content to below 8%, thereby preventing microbial growth and preserving the stability of heat-sensitive polyphenolic compounds. Pulverization of the dried material increased the surface area available for solvent penetration, resulting in improved extraction efficiency. The hydroalcoholic extraction system utilizing ethanol and water in a 70:30 ratio demonstrated effective recovery of punicalagins and other polyphenols from the peel matrix.

Following extraction, filtration and vacuum concentration successfully removed insoluble materials and reduced solvent volume without exposing the extract to excessive heat. Maintaining the concentration temperature below 55°C was critical for preserving the structural integrity of punicalagins.

Resin purification using adsorption resins such as AB-8, XAD-16, and Diaion HP20 significantly enhanced the concentration of target compounds by selectively removing unwanted impurities and low-molecular-weight constituents.

Further purification through chromatographic techniques, including Preparative HPLC, Flash Chromatography, and MPLC, enabled efficient separation of α-punicalagin and β-punicalagin from other phenolic compounds. The final crystallization process produced a highly purified product with excellent physicochemical characteristics and minimal contamination. HPLC analysis confirmed that the combined concentration of α-punicalagin and β-punicalagin exceeded 98%, meeting the target specification for pharmaceutical and nutraceutical applications.

The physicochemical evaluation demonstrated that the extract complied with established quality standards. The product exhibited acceptable moisture levels, low residual solvent content, controlled heavy metal concentrations, and satisfactory microbiological quality. These findings indicate that the manufacturing process is capable of consistently producing a safe, stable, and standardized extract.

The high purity achieved in the final product is particularly significant because punicalagins are responsible for many of the biological activities associated with pomegranate peel. Numerous studies have demonstrated that punicalagins possess exceptional antioxidant activity due to their ability to neutralize reactive oxygen species and prevent oxidative damage to cellular components. The high concentration of these compounds in the standardized extract enhances its potential effectiveness in nutraceutical and pharmaceutical formulations.

In addition to antioxidant properties, the extract exhibited characteristics associated with strong antimicrobial and anti-inflammatory potential. The purification process concentrated the active polyphenols while minimizing unwanted constituents, thereby improving the overall functional quality of the extract. The resulting product is suitable for applications targeting cardiovascular health, immune support, healthy aging, metabolic wellness, and cellular protection.

Overall, the results demonstrate that the combination of hydroalcoholic extraction, adsorption resin purification, chromatographic refinement, and controlled crystallization provides an effective strategy for producing high-purity Punicalagins 98%. The manufacturing process is scalable, reproducible, and capable of delivering a premium-quality ingredient suitable for use in pharmaceutical, nutraceutical, functional food, and cosmetic industries.

## Benefits and Mechanism of Action of Punicalagins

Punicalagins are the major bioactive compounds present in pomegranate peel and are recognized for their exceptional antioxidant, anti-inflammatory, antimicrobial, and protective properties. These water-soluble ellagitannins contribute significantly to the therapeutic potential of pomegranate

extract and have been extensively studied for their role in promoting overall health and preventing chronic diseases.

#### Antioxidant Benefits

One of the most important biological activities of punicalagins is their powerful antioxidant capacity. Their polyphenolic structure enables efficient scavenging of free radicals and protection against oxidative stress.

- Neutralizes reactive oxygen species (ROS) and reactive nitrogen species (RNS).
- Protects lipids, proteins, and DNA from oxidative damage.
- Reduces cellular injury caused by free radicals.
- Supports endogenous antioxidant enzymes such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx).
- Helps slow age-related oxidative damage and cellular degeneration.

These antioxidant properties contribute to the prevention of various chronic disorders associated with oxidative stress.

#### Anti-Inflammatory Benefits

Chronic inflammation plays a significant role in the development of cardiovascular diseases, metabolic disorders, and degenerative conditions. Punicalagins help regulate inflammatory responses through multiple pathways.

- Inhibits the production of pro-inflammatory cytokines.
- Suppresses activation of the NF- $\kappa$ B signaling pathway.
- Reduces inflammatory mediator release.
- Helps protect tissues from chronic inflammatory damage.
- Supports healthy immune system regulation.

By controlling inflammation, punicalagins contribute to long-term tissue protection and overall wellness.

#### Cardiovascular Benefits

Punicalagins have demonstrated significant cardioprotective effects through their antioxidant and anti-inflammatory actions.

- Protects blood vessels from oxidative stress.
- Reduces oxidation of low-density lipoprotein (LDL) cholesterol.
- Supports healthy endothelial function.
- Promotes normal blood circulation.
- Helps maintain cardiovascular health.

These effects may contribute to reducing risk factors associated with cardiovascular diseases.

#### Antimicrobial Benefits

Punicalagins exhibit broad-spectrum antimicrobial activity against numerous pathogenic microorganisms.

- Inhibits the growth of Gram-positive and Gram-negative bacteria.
- Demonstrates antifungal activity against several fungal species.

- May interfere with viral attachment and replication.
- Prevents microbial colonization and biofilm formation.
- Supports natural defense mechanisms against infections.

These antimicrobial properties increase the value of punicalagins in pharmaceutical, nutraceutical, and food applications.

#### Cellular Protection and Healthy Aging

The ability of punicalagins to protect cellular structures contributes to healthy aging and maintenance of physiological function.

- Protects cellular membranes from oxidative injury.
- Preserves mitochondrial function and energy production.
- Supports tissue repair and regeneration.
- Reduces accumulation of oxidative damage over time.
- Promotes overall cellular health and longevity.

#### Mechanism of Action

The biological effects of punicalagins are mediated through multiple interconnected molecular mechanisms that begin following oral consumption.

##### Step 1: Hydrolysis and Metabolism

- Punicalagins are hydrolyzed in the gastrointestinal tract.
- Ellagic acid is released from the punicalagin molecule.
- Gut microbiota convert ellagic acid into bioactive urolithins.

These metabolites are absorbed and distributed throughout the body, where they exert systemic biological effects.

##### Step 2: Antioxidant Action

- Directly scavenges free radicals.
- Inhibits oxidative chain reactions.
- Prevents lipid peroxidation.
- Maintains cellular redox balance.

This mechanism helps reduce oxidative stress and cellular damage.

##### Step 3: Anti-Inflammatory Action

- Suppresses inflammatory signaling pathways.
- Inhibits activation of NF- $\kappa$ B.
- Reduces cytokine production.
- Limits inflammatory tissue damage.

These effects contribute to protection against chronic inflammatory conditions.

##### Step 4: Cardiovascular Protection

- Protects endothelial cells.
- Improves vascular function.
- Reduces oxidative modification of lipoproteins.
- Supports healthy blood vessel integrity.

This mechanism promotes cardiovascular wellness and circulatory health.

### Step 5: Antimicrobial Action

- Disrupts microbial cell membranes.
- Alters membrane permeability.
- Interferes with microbial enzyme systems.
- Inhibits microbial growth and survival.

These actions contribute to the antimicrobial properties of pomegranate extract.

Through these combined mechanisms, Punicalagins 98% provide comprehensive antioxidant, anti-inflammatory, antimicrobial, and protective effects that support overall health, cellular integrity, and disease prevention.

### CONCLUSION

Punicalagins are the principal bioactive ellagitannins present in pomegranate (*Punica granatum* L.) peel and are responsible for many of the plant's scientifically recognized health benefits. The present study demonstrated an effective industrial manufacturing process for the production of high-purity pomegranate extract standardized to 98% punicalagins. The combination of controlled drying, hydroalcoholic extraction, filtration, vacuum concentration, resin-based purification, chromatographic refinement, and crystallization enabled the successful isolation and enrichment of punicalagins while preserving their biological activity and chemical stability.

Physicochemical evaluation confirmed that the final product met established quality specifications with respect to purity, moisture content, microbial limits, residual solvents, and heavy metal concentrations. HPLC analysis verified the presence of  $\alpha$ -punicalagin and  $\beta$ -punicalagin as the major active constituents and confirmed a total punicalagin content of not less than 98%, ensuring product standardization and batch-to-batch consistency.

The high concentration of punicalagins contributes to the extract's exceptional antioxidant, anti-inflammatory, antimicrobial, and protective properties. Through their ability to neutralize free radicals, regulate inflammatory pathways, support cardiovascular function, and protect cellular structures, punicalagins offer significant potential for nutraceutical, pharmaceutical, functional food, and cosmetic applications. Furthermore, their conversion to bioactive metabolites such as ellagic acid and urolithins enhances their systemic biological effects and therapeutic value.

Overall, the manufacturing process described in this study provides a scalable, reproducible, and commercially viable approach for producing pharmaceutical-grade Punicalagins 98%. The resulting extract represents a high-value natural ingredient with broad applications in health promotion, disease prevention, and advanced nutritional supplementation. Continued research into the biological activities and clinical benefits of punicalagins is expected to further expand their role in evidence-based healthcare and functional product development.

### REFERENCES

1. Afaq, F., Malik, A., Syed, D., Maes, D., Matsui, M. S., & Mukhtar, H. (2005). Pomegranate fruit extract modulates UV-B-mediated phosphorylation of MAPK and activation of NF- $\kappa$ B in normal human epidermal keratinocytes. *Photochemistry and Photobiology*, 81(1), 38–45.
2. Bialonska, D., Ramnani, P., Kasimsetty, S. G., Muntha, K. R., Gibson, G. R., Ferreira, D., & Gross, H. B. (2010). The influence of pomegranate by-product and punicalagins on selected groups of human intestinal microbiota. *International Journal of Food Microbiology*, 140(2–3), 175–182.
3. Cerda, B., Llorach, R., Ceron, J. J., Espin, J. C., & Tomas-Barberan, F. A. (2003). Evaluation of the bioavailability and metabolism in the rat of punicalagin, an antioxidant polyphenol from pomegranate juice. *European Journal of Nutrition*, 42(1), 18–28.
4. Gil, M. I., Tomas-Barberan, F. A., Hess-Pierce, B., Holcroft, D. M., & Kader, A. A. (2000). Antioxidant activity of pomegranate juice and its relationship with phenolic composition and processing. *Journal of Agricultural and Food Chemistry*, 48(10), 4581–4589.
5. Fischer, U. A., Carle, R., & Kammerer, D. R. (2011). Identification and quantification of phenolic compounds from pomegranate peel, mesocarp, aril and differently produced juices. *Food Chemistry*, 127(2), 807–821.
6. Ismail, T., Sestili, P., & Akhtar, S. (2012). Pomegranate peel and fruit extracts: A review of potential anti-inflammatory and anti-infective effects. *Journal of Ethnopharmacology*, 143(2), 397–405.
7. Jurenka, J. S. (2008). Therapeutic applications of pomegranate (*Punica granatum* L.): A review. *Alternative Medicine Review*, 13(2), 128–144.
8. Kaderides, K., Papaoikonomou, L., Serafim, M., & Goula, A. M. (2015). Microwave-assisted extraction of phenolics from pomegranate peels. *Food Chemistry*, 176, 405–411.
9. Kulkarni, A. P., Aradhya, S. M., & Divakar, S. (2004). Isolation and identification of a radical scavenging antioxidant from pomegranate peel. *Journal of the Science of Food and Agriculture*, 84(8), 892–896.
10. Lansky, E. P., & Newman, R. A. (2007). Punica granatum (pomegranate) and its potential for prevention and treatment of inflammation and cancer. *Journal of Ethnopharmacology*, 109(2), 177–206.
11. Li, Y., Guo, C., Yang, J., Wei, J., Xu, J., & Cheng, S. (2006). Evaluation of antioxidant properties of pomegranate peel extract. *Food Chemistry*, 96(2), 254–260.
12. Mena, P., Calani, L., Dall'Asta, C., Galaverna, G., Garcia-Viguera, C., Bruni, R., Crozier, A., & Del

- Rio, D. (2012). Rapid and comprehensive evaluation of pomegranate polyphenols by UHPLC-MS. *Journal of Agricultural and Food Chemistry*, 60(23), 5600–5607.
13. Negi, P. S., & Jayaprakasha, G. K. (2003). Antioxidant and antibacterial activities of pomegranate peel extracts. *Food Microbiology*, 20(4), 393–397.
14. Ozgen, M., Durgac, C., Serce, S., & Kaya, C. (2008). Chemical and antioxidant properties of pomegranate cultivars grown in the Mediterranean region. *Food Chemistry*, 111(3), 703–706.
15. Pagliarulo, C., De Vito, V., Picariello, G., Colicchio, R., Pastore, G., Salvatore, P., & Volpe, M. G. (2016). Inhibitory effect of pomegranate peel extract on bacterial growth and biofilm formation. *BMC Complementary and Alternative Medicine*, 16(1), 1–10.
16. Seeram, N. P., Adams, L. S., Henning, S. M., Niu, Y., Zhang, Y., Nair, M. G., & Heber, D. (2005). In vitro antiproliferative, apoptotic and antioxidant activities of punicalagin. *Journal of Agricultural and Food Chemistry*, 53(4), 1496–1503.
17. Seeram, N. P., Lee, R., Hardy, M., & Heber, D. (2005). Rapid large-scale purification of ellagitannins from pomegranate husk. *Journal of Agricultural and Food Chemistry*, 53(15), 6578–6585.
18. Singh, R. P., Chidambara Murthy, K. N., & Jayaprakasha, G. K. (2002). Studies on antioxidant activity of pomegranate peel and seed extracts. *Journal of Agricultural and Food Chemistry*, 50(1), 81–86.
19. Tomas-Barberan, F. A., & Espin, J. C. (2001). Phenolic compounds and related enzymes as determinants of quality in fruits and vegetables. *Journal of the Science of Food and Agriculture*, 81(9), 853–876.
20. Viuda-Martos, M., Fernandez-Lopez, J., & Perez-Alvarez, J. A. (2010). Pomegranate and its many functional components as related to human health. *Comprehensive Reviews in Food Science and Food Safety*, 9(6), 635–654.
21. Wang, R., Ding, Y., Liu, R., Xiang, L., Du, L., & Piao, X. (2010). Purification and identification of punicalagins from pomegranate husk. *Food Chemistry*, 123(1), 221–226.
22. Yuan, T., Ma, H., Liu, W., Niesen, D. B., Shah, N., Crews, R., Rose, K. N., Vatter, D. A., & Seeram, N. P. (2016). Pomegranate's neuroprotective and antioxidant properties. *Food Chemistry*, 194, 156–165.
23. Zhang, L., Gao, Y., Zhang, Y., Liu, J., Yu, J., & Zhao, J. (2014). Effects of extraction methods on polyphenol yield and antioxidant activity of pomegranate peel extract. *Food Analytical Methods*, 7(5), 1095–1102.
24. Elfalleh, W., Hannachi, H., Tlili, N., Yahia, Y., Nasri, N., & Ferchichi, A. (2012). Total phenolic contents and antioxidant activities of pomegranate peel extracts. *Industrial Crops and Products*, 35(1), 287–293.
25. Akhtar, S., Ismail, T., Fraternal, D., & Sestili, P. (2015). Pomegranate peel and peel extracts: Chemistry and health benefits. *Food Bioscience*, 9, 11–20.
26. Hayouni, E. A., Miled, K., Boubaker, S., Bellasfar, Z., Abedrabba, M., Iwaski, H., Oku, H., Matsui, T., Limam, F., & Hamdi, M. (2011). Hydroalcoholic extraction and biological activities of pomegranate peel. *Food Chemistry*, 125(2), 578–584.
27. Kaderides, K., Goula, A. M., & Adamopoulos, K. G. (2015). Process optimization for extraction of bioactive compounds from pomegranate peel. *Waste and Biomass Valorization*, 6(6), 957–965.
28. Cam, M., Hisil, Y., & Durmaz, G. (2009). Classification of pomegranate juices based on antioxidant capacity and phenolic composition. *Food Chemistry*, 112(3), 721–726.
29. Tezcan, F., Gultekin-Ozguven, M., Diken, T., Ozcelik, B., & Erim, F. B. (2009). Antioxidant activity and total phenolic content of pomegranate extracts. *Food Chemistry*, 115(3), 873–877.
30. Johanningsmeier, S. D., Harris, G. K., & Klevorn, C. M. (2011). Metabolic effects and bioavailability of pomegranate polyphenols and punicalagins. *Journal of Medicinal Food*, 14(11), 1261–1268.